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## Non-chromatographic hydride generation atomic spectrometric techniques for the speciation analysis of arsenic, antimony, selenium, and tellurium in water samples—a review

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## Non-chromatographic hydride generation atomic spectrometric techniques for the speciation analysis of arsenic, antimony, selenium, and tellurium in water samples—a review

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Hydride generation (HG) coupled with AAS, ICP–AES, and AFS techniques for the speciation analysis of As, Sb, Se, and Te in environmental water samples is reviewed. Careful control of experimental conditions, offline/online sample pretreatment methods employing batch, continuous and flow-injection techniques, and cryogenic trapping of hydrides enable the determination of various species of hydride-forming elements without the use of chromatographic separation. Other non-chromatographic approaches include solvent extraction, ion exchange, and selective retention by microorganisms. Sample pretreatment, pH dependency of HG, and control of NaBH<sub>4</sub>/HCl concentration facilitate the determination of As(III), As(V), monomethylarsonate (MMA), and dimethylarsinate (DMA) species. Inorganic species of arsenic are dominant in terrestrial waters, whereas inorganic and methylated species are reported in seawater. Selenium and tellurium speciation analysis is based on the hydrides generation only from the tetravalent state. Se(IV) and Se(VI) are the inorganic selenium species mostly reported in environmental samples, whereas speciation of tellurium is rarely reported. Antimony speciation analysis is based on the slow kinetics of hydride formation from the pentavalent state and is mainly reported in seawater samples.

Keywords: Non-chromatographic hydride generation techniques; Speciation analysis; Arsenic; Antimony; Selenium; Tellurium

## 1. Introduction

Assessment of environmental risk caused by metalloids such as As, Sb, Se, and Te is widely recognized to be dependent upon speciation. According to the latest IUPAC [1] guidelines, the term 'speciation analysis' refers to analytical activities of identifying and/ or measuring the quantities of one or more chemical species in a sample. The term chemical species refers to the specific form of an element, defined as molecular, complex, or nuclear structure or oxidation state. It is now well established that information about the speciation of an element is required for understanding the

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reaction of many elements. For example, the inorganic forms of arsenic and selenium exhibit different toxicities, i.e.  $As(III)$  is more toxic than  $As(V)$ , and  $Se(IV)$  is more toxic than Se(VI). However, methylated forms of these elements are relatively less toxic than their inorganic forms, and organic forms of arsenic, like arsenobetaine and arsenocholine, are virtually non-toxic. In addition, knowledge of speciation could help to explain the mobility, bioavailability, storage, and retention of elemental species in different environmental matrices including the human body.

Speciation analysis usually involves two steps: (1) separation of different forms and (2) their subsequent quantification. A wide variety of techniques, based on liquid chromatography interfaced with atomic spectrometric techniques, known as hyphenated techniques, i.e. HPLC–HGAAS [2], HPLC–HGAFS [3], HPLC–HG–ICP–mass spectrometry (MS) [4], etc., have been proposed for the speciation of hydride-forming elements and reviewed recently [5]. The primary advantage of this approach is the unequivocal species separation and specific online detection. Unfortunately, most of these techniques have significant disadvantages [6]: increased complexity, co-elution of species of the same element [7], problems associated with the stability of plasma due to the use of organic solvents, and interference arising from polyatomic ions in conventional ICP–MS [8]. Additionally, these types of hyphenated instruments are not standard equipment in most laboratories engaged in water-quality monitoring. Further considerations to be taken into account are the acquisition and running costs, analysis time, and ease of handling, especially for routine analysis.

HG coupled with atomic spectrometric techniques is a sensitive analytical tool for the determination of trace levels of As, Sb, Se, and Te. The popularity of HG, nearly 30 years since its introduction as an analytical technique, is still increasing because of its ability in differentiating different oxidation states of these elements by simple procedures, without the use of chromatographic separation. By applying suitable separation techniques, elemental species which form the corresponding hydrides can be quantitated. The separation techniques include selective hydride formation based on pH adjustment, buffer, reaction media, matrix modification, complexation-extraction, selective retention, etc. Another approach is to collect the hydrides in a cold trap and their sequential release to the detection system. In addition, a variety of modern automated flow-injection (FI) and continuous flow (CF) approaches utilizing efficient hyphenations with online sample/analyte treatment (pre- or post-HG or both) such as sample introduction, decomposition, prereduction, ultraviolet photo oxidation, microwave decomposition, enrichment, solid-phase extraction, etc. constitute the various approaches for elemental speciation.

#### 2. Previous reviews

We refer readers to the references listed in table 1 for recent general reviews focused on speciation analysis of metalloids by HG coupled with various detectors. The other complementary reviews include new developments, extraction techniques, stability studies, etc. More information on speciation analysis of arsenic, antimony, selenium, and tellurium can be found in the articles listed in tables 2 and 3.





Table 1. General reviews on HG techniques in speciation analysis and other complementary reviews.<sup>4</sup> Table 1. General reviews on HG techniques in speciation analysis and other complementary reviews.<sup>a</sup>  $(Continued) % \begin{minipage}[b]{0.5\linewidth} \centering \centerline{\includegraphics[width=0.5\linewidth]{images/STM100020.jpg} \centerline{\includegraphics[width=0.5\linewidth]{images/STM100020.jpg} \centerline{\includegraphics[width=0.5\linewidth]{images/STM100020.jpg} \centerline{\includegraphics[width=0.5\linewidth]{images/STM100020.jpg} \centerline{\includegraphics[width=0.5\linewidth]{images/STM100020.jpg} \centerline{\includegraphics[width=0.5\linewidth]{images/STM100020.jpg} \centerline{\includegraphics[width=0.5\linewidth]{images/STM100020.jpg} \centerline{\includegraphics[width$ (Continued)

analysis of As, Sb, Se, and other elements



Table 1. Continued. Table 1. Continued.

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Table 2. Arsenic speciation analysis reviews. Table 2. Arsenic speciation analysis reviews.

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in environmental water samples



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Table 3. Antimony, selenium, and tellurium speciation analysis reviews. J.  $\ddot{\phantom{a}}$ J. intio J.  $\overline{d}$  telling J.  $\frac{1}{2}$  $\frac{1}{2}$  $\lambda$ <sup>+</sup>  $\ddot{\phantom{0}}$  $TaMa$ 

#### 3. Aims and scope

Although this review deals primarily with recent developments in HG techniques based on non-chromatographic methods, it is not aimed solely at an analytical audience but is intended for all scientists with an interest in speciation analysis. It is hoped that the information will be accessible to, and useful for, scientists in interdisciplinary fields such as environmental science and toxicology. The aim of this review is to highlight the developments, achievements, current state, analytical scope, performance characteristics, advantages, limitations, and prospects of HG coupled with various atomic spectrometric detectors for the speciation analysis of As, Sb, Se, and Te in environmental water samples. Emphasis is placed on the methods reported after the year 2000, although some previous publications are considered for performance evaluation. As we focus on simple procedures based on HG, hyphenated techniques involving MS are not discussed.

The concentrations of several arsenic, antimony, and selenium species in different environmental water samples are reported in tables 4–6.

#### 4. Arsenic speciation analysis

#### 4.1 Development of HG as a speciation technique

In 1973, Braman and Foreback [87] reported the first method of arsenic speciation, i.e. arsenite, arsenate, monomethylarsonate (MMA), and dimethylarsinate (DMA) based on selective HG and sequential volatilization. The evolved arsenicals were collected in a liquid nitrogen trap, warmed, and sequentially released according to their boiling point (arsine  $-55^{\circ}$ C, methylarsine  $2^{\circ}$ C, dimethylarsine  $55^{\circ}$ C) and determined by the DC arc emission method. The work of Braman and Foreback may be seen as heralding in the field of speciation analysis, in particular, arsenic speciation. In subsequent years, research in speciation analysis grew steadily, exploiting the pH dependency of HG. In the last two decades, interest in arsenic speciation has been heightened by toxicological issues, in particular the carcinogenic effects of inorganic arsenic in drinking water and the worldwide human-health implications [88, 89]. In addition, arsenic is also attracting renewed clinical interest and is currently posting some remarkable success in the treatment of a certain type of leukemia [90].

#### 4.2 Speciation analysis based on selective HG

The reaction between tetrahydroborate (THB) and an oxyion in solution is sensitive to pH, and for the reaction to proceed rapidly, the analyte species must be fully protonated. As  $pK_1$  for arsenic acid is 2.3, the reaction must therefore be carried out at very low pH, i.e.  $1-2 \text{ mol L}^{-1}$  HCl is generally employed. Arsenite (pK<sub>1</sub> 9.2), on the other hand, is protonated under natural water conditions, and reacts with THB under mild acidic conditions. Thus, differentiation of  $As(III)$  and  $As(V)$  could be achieved simply by exploiting the pH dependency of THB reduction.



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> Coppt: co-precipitation.<br>
> PAverage of six samples and As<sub>T</sub> includes 81.1 µg L<sup>-1</sup> of particulate As.<br>
> "Average of three samples.<br>
> nd: not detected. cAverage of three samples. nd: not detected.

Coppt: co-precipitation.

<sup>b</sup>Average of six samples and As<sub>T</sub> includes 81.1 µg  $L^{-1}$  of particulate As.



Concentration of selemium species in environmental samples. Table 5. Concentration of selenium species in environmental samples.

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nd: not detected.



nd: not detected.

Table 6. Concentration of antimony species in environmental water samples.  $\cdot$  $ntal<sub>1</sub>$ ÷ J, .9  $\frac{1}{2}$  $\epsilon$  $\frac{1}{2}$ ŧ Č  $\check{\phantom{a}}$  $Tahle$ 

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Aggett and Aspell [91] first reported the determination of As(III) and total As by determining As(III) at a pH of 4–5 and total As with  $5 \text{ mol} L^{-1}$  HCl. Nakashima [92] reported the determination of As(III) in presence of As(V) at  $0.25 \text{ mol} L^{-1}$  HCl with  $3-5$  mg mL<sup>-1</sup> of Zr(IV) and 2% KI. Total As was determined in  $2 \text{ mol} L^{-1}$  HCl containing  $1\%$  KI. The Zr(IV), along with  $2\%$  KI, selectively suppressed the signal of As(V) in As(III) determination. Based on the pH dependency of arsine generation, several workers reported the speciation of inorganic arsenic [93–96]. Soon it was realized that methylated species also form their corresponding methylatedarsines and could be determined in the same way as their inorganic forms [97].

Hinners [98] pointed out the major drawback of this procedure. He reported that if methylated arsenic is present, it could also contribute to the signal of As(III) and total As, and suggested that buffer solutions having a sufficient buffer capacity could minimize this error, and species-specific standards should be used for calibration. Lopez *et al.* [99] determined As(III) in citrate buffer and total As in 6 mol  $L^{-1}$  HCl without prereduction. They [99] reported that MMA and DMA interfered in the determination of As(III) and total As. Other workers suggested that not only does the pH affect the speciation but other factors such as kinetics and complexation are also involved [99, 100]. Several authors [101–104] reported the use of acetate and citrate buffers for As(III) determination at pH  $\approx$  5 and total arsenic at higher acidities, i.e. 1–5 mol L<sup>-1</sup> acid with KI/KI-ascorbic acid prereduction. However, all these studies continued to report only the inorganic As speciation analysis.

Anderson *et al.* [105] reported a selective reduction method to speciate inorganic and methylated forms using different buffer matrices. One important finding of this work was that in mercaptoacetic acid medium, the four arsenic species (As(III), As(V), MMA, and DMA) showed similar response profiles. They proposed that the sulfurcontaining ligand must play a significant role in effecting rapid hydride generation, probably, by altering reaction mechanism. Brindle et al. [106] and Chen et al. [107] introduced the use of  $L$ -cysteine as a prereductant for  $As(V)$  and reported that it reduces the acid concentration required for hydride formation and minimizes interferences form transition metals [108]. The use of L-cysteine gained popularity as a prereductant, and many procedures have included its use for As speciation analysis. Following the method of Anderson et al. [105], several articles were published on arsenic speciation analysis utilizing different acids, buffers, and redox media to determine inorganic [109–111] and methylated species [61–64, 112–117]. Typical procedures include the work of Torralba *et al.* [112, 113], Yano *et al.* [114], Rude and Puchelt [115], Quinaia *et al.* [116], and Shraim *et al.* [64, 117]. Shrim *et al.* [64] proposed the use of  $HClO<sub>4</sub>$  and claimed that it is the only acid that facilitates the determination of total inorganic As without the interference from methylated species.

Selective hydride generation offers a simple and inexpensive practical tool for As speciation analysis. However, there are several inherent disadvantages: long reaction times, allow sample throughput, and an increased use of chemicals besides the strict control of reaction conditions are required.

Another approach for As speciation analysis is based on the use of appropriate concentration of acid and THB for selective generation of hydrides. However, this approach is limited to the determination of inorganic As species only. Most procedures use a low concentration of acid/THB to generate  $\text{AsH}_3$  from As(III) and a higher concentration of acid/THB for total As determination. This approach is based on the redox property of THB, which acts as a reductant and hydride source.

Narsito and Agterdenbos [100] observed a significant difference in the reduction efficiency between As(III) and As(V) at THB concentrations  $\leq 1\%$  and strongly acidic media. They suggested that As(III) is converted to  $AsH<sub>3</sub>$  more efficiently than As(V) under these conditions. Borho and Wilderer [118] reported the use of low THB concentrations under highly acidic conditions, i.e.  $pH < 0$  to generate AsH<sub>3</sub> only from As(III). This is particularly useful for acid-preserved samples. Muller [119] reported the selective  $\text{AsH}_3$  formation from As(III) using a low THB concentration. Sigrist and Beldomenico [49] reported a similar procedure using the FI–HGAAS system for samples containing  $\leq 1 \mu g L^{-1}$  of methylated As.

This approach is found to introduce serious errors, if methylated arsenicals are present. In addition, the use of low concentrations of THB decreases the sensitivity, and so the limit of detection (LOD) would be affected. In contrast, this approach offers an extended linear working range.

Other approaches for selective determination of As species include selective extraction and co-precipitation techniques [10]. These techniques allow the separation of one or more species of interest and subsequent HG. One important advantage of this approach is that interference from methylated species is greatly minimized. Generally, ammonium pyrrolidinedithiocarbamate (APDC) [120–124] or ammonium sec-butyldithiophosphate [125] is used to selectively complex As(III) at pH 3.5–5.5.

Yu et al. [85] reported selective retention of As(III) on thiol cotton (cotton treated with thioglycolic acid), which was eluted by hot HCl. As(V) was determined after KI–thiourea prereduction. Bombach et al. [126] reported the selective retention of As(V) by an anion-exchange column for on-site species separation. Okumura *et al.* [127] reported a zirconium loaded sorbent for the retention of As(III) and As(V), which were selectively eluted with  $0.5 \text{ mol L}^{-1}$  of HCl and NaOH, respectively. Chatterjee *et al.* [128] described a method to specifically extract As(III) with sodium diethyldithiocarbamate at pH 5.5 in  $\text{CCI}_4$  and subsequent determination by HGAAS. Samanta *et al.* [56] followed the same procedure to speciate inorganic arsenic. Dapaah and Ayame [129] extracted As(III) with zinc hexamethylenedithiocarbamate complex in 2,6-dimethyl-4-heptanone. The organic phase was back-extracted with Cu(II) and determined by FI–HGAAS. Van Elteren et al. [130] determined As(V) and As(III) by co-precipitating As(V) as molybdate complex with tetraphenylphosphoniumchloride and As(III) with dibenzyldithiocarbamate.

## 4.3 HG–CT techniques

The discovery of dissolved methylated arsenic species in the marine environment and most of the subsequent work which has been carried out to understand the presence and significance of related antimony, selenium, and tellurium species has resulted from the development of cryogenic-trap hydride methods. Cryogenic trapping of the volatile hydrides at liquid- $N_2$  temperatures results in a focusing of the products as a narrow plug of material which can then be separated either by a distillation mechanism using glass beads or a chromatographic material packed column. The collected arsines differ significantly in their boiling points (b.p. AsH<sub>3</sub> -55°C, CH<sub>3</sub>AsH<sub>2</sub> 2°C and (CH<sub>3</sub>)<sub>2</sub>AsH 55-C) and can be sequentially evolved from the trap by heating. With a glass-bead trap, this is simply achieved by removing it from liquid  $N_2$ , but, if a chromatographic material packing is involved, it must be heated electrically. The arsines are then detected with different atomic spectrometric methods.

The first reported As speciation method by Braman and Foreback [87] utilized the trap-speciation method. They utilized a 'U' tube packed with glass beads. Subsequently, several improvements to this trap system have been reported [131, 132]. Andreae [133] described a method with considerable improvement in the system with air– $H_2$  flame AAS and attained LODs in  $ng L^{-1}$  levels. The glass beads were replaced by silanized glass wools, which gave quantitative recovery compared with silanized glass beads (80%). Howard and Arbab-Zavar [134] utilized a continuous flow hydride generator with CT-QFAAS detection. This greatly improved the performance and minimized transition metal interferences. Van Cleuvenbergen et al. [135] determined As(III) in Tris buffer (pH 7.5) and total arsenic at pH 0.8. Anderson et al. [136] used phthalate buffer for As(III) reduction and oxalic acid for determining other species. Ryu et al. [137] determined As(III) at pH 6, and total arsenic and methylated species were determined at pH 1.5 using oxalic acid.

Howard and Comber [138] obtained an improved sensitivity by changing the CF manifold geomentry, etching the glass beads with HF instead of silanization used previously, and obtained an LOD of 19–61 ng  $L^{-1}$  using 1 mL of sample. Michel *et al.* [139] reported an improvement in the technique with the use of  $H_2SO_4$  instead of HCl and progressive addition of THB. According to these authors [139], this resulted in a smoother and efficient response, and an LOD of  $10 \text{ ng } L^{-1}$  was reported. A further remarkable improvement in species separation was reported by van Elteren [60], by coprecipitation of As(III) with sodium dibenzyldithiocarbamate and As(V) with tetraphenylephosphoniumchloride as molybdate complex and determined separately. The methylated arsines were passed through a novel perma-pure filter, collected in a stainless steel trap, and determined by QFAAS. The advantage of this trap is faster heating and cooling, thus yielding a better species separation. The reported LOD was  $0.16 \mu g L^{-1}$ . Burguera et al. [140] reported arsine generation by Fleitmann reaction, i.e. using Al–NaOH for inorganic arsenic speciation analysis. The advantage of this procedure is the use of low-cost, stable, and easily stored reagents compared with the THB/HCl system.

Howard and Salou [141] applied cysteine pretreatment method, which led to a sensitivity enhancement and better interference control, and attained an LOD of 50 ng  $L^{-1}$ . In another study [142], they investigated various prereductants for use in HG–CT–QFAAS and reported that thiglycolic acid also produced the same effect as that of cysteine. Burguera et al. [57] reported a fully automatic FI–HG–CT–AAS. The trap was made of PTFE coil and heated by microwave radiation. The reported LODs were 20, 24, 35, and 60 ng  $L^{-1}$  for As(III), As(V), MMA, and DMA, respectively. Molenat et al. [62] used phosphate buffer (pH 6.8) for As(III) and oxalic acid for  $As(III) + As(V)$ , MMA, DMA, and trimethylarsine oxide (TMAO) determination. Narvaez *et al.* [48] reported the online removal of copper as iminodiacetate complex using a microcolumn prior to HG. As(III) was determined selectively using acetate buffer (pH 4.5), and  $As(III) + As(V)$ , MMA, and DMA were determined at pH 1.

Another remarkable achievement in the CT technique was the determination of methylated species of As(III) and As(V). Hasegava *et al.* [143] reported a solvent extraction procedure using the diethylammonium diethyldithiocarbamate (DDDC) procedure to determine the methylated trivalent arsenic species and methylated pentavalent arsenic species, in addition to inorganic species. In another study [144], they

described a method to determine the non-hydride fraction, in addition to the hydride reactive species. According to them, the non-hydride fraction may be divided into ultraviolet (UV)-labile and UV-inert fractions based on their conversion into hydride active species. Cabon and Cabon [145] determined hydride active species by alkaline persulfate digestion, whereas the same matrix after UV irradiation converted the inactive species to hydride active species and quantified by FI–CT–HGAAS. Recently, Hsiung and Wang [146] reported a novel packed cold finger trap (PCFT) packed with 10% OV-101 on chromosorb W-HP. This led to a better performance in terms of species separation than the 'U' trap. LODs of 0.047, 0.042, 0.0045, and 0.0063  $\mu$ g L<sup>-1</sup> were reported for As(III), As(V), MMA, and DMA, respectively.

Cryogenic trapping methods offer efficient species separation and improved LOD to  $ng L<sup>-1</sup>$  levels. The methods reported earlier used glass beads as packing material, which led to incomplete species recovery and separation. These drawbacks are minimized in the chromatographic material filled traps. However, the complexity of the system as a whole makes this method less attractive for routine monitoring.

#### 4.4 FI–HGAAS techniques

The introduction of FI principles and techniques into HG methods in AAS, pioneered by Astrom's [147] work in 1982, established an important milestone in the development of HGAAS. The topic has been reviewed by several authors [12, 13, 16]. FIA offers many advantages; the main features include a shorter analysis time, less consumption of sample/reagents, reduced human intervention, better sensitivity and precision, and increased tolerance to interference. The flexibility of this approach to include online separation techniques based on ion exchange, complexation/solvent extraction, solidphase extraction (SPE), oxidation/reduction, etc. enabled its applicability as a powerful tool in speciation analysis.

Differentiation of  $As(III)$  and  $As(V)$  was performed by online pH-dependent arsine generation either utilizing varying acid/THB concentration or using appropriate buffers. Determination of As(III) and total As after online reduction using KI/KI–ascorbic acid [148–150], thiosulfate [124], and L-cysteine [151–154] was reported by many workers. Stopped flow techniques were developed to allow the reduction of As(III) species utilizing KI [155] and  $4 \text{ mol} L^{-1}$  HCl [156]. The use of L-cysteine avoids the problems faced with other reductants such as KI and HCl. Lee et al. [157] showed that L-cysteine is an effective pre-derivatizing agent for inorganic as well as methylated arsenic species. Carrero et al. [158] studied the pre-derivatization of As species with L-cysteine and reported that at low acid concentrations, arsine is formed by the action of THB with analyte–thiolate complex, and at high concentration of acid, arsine is produced by a nascent hydrogen mechanism. Nielsen and Hansen [58] reported an online reduction of As(V) with KI–ascorbic acid in a heated knotted reactor. Torralba et al. [159] determined As(III) selectively in citrate buffer and total arsenic with 6 mol  $L^{-1}$  HCl. Online co-precipitation with La(OH)<sub>3</sub> in a knotted reactor for selective As(III) determination was also reported [160], with an LOD of  $3 \text{ ng } L^{-1}$ .

In contrast to the online reduction, Coelho et al. [52] used  $1 \text{ mol } L^{-1}$  of HCl with 0.1% THB for selective As(III) determination and 3% THB for total As determination. This avoids the prereduction step reported by many authors [148–153]. The use of a low

THB concentration, i.e.  $0.035\%$  with  $2 \text{ mol L}^{-1}$  HCl for As(III) and total As after KI reduction, was reported by Sigrisit and Beldomenico [49].

Online pre-concentration of As(III) on activated alumina and total As after prereduction with L-cysteine was also reported [161]. Simultaneous online pre-concentration using an anion-exchange column was reported by Narcise et al. [45], in which, the column is selective to As(V) at neutral pH and total As at pH  $> 12$ . Interference from methylated species was removed by cation exchange before selective adsorption. Anthemidis et al. [46] reported a sequential injection system, in which, 0.5% THB with  $1.5 \text{ mol L}^{-1}$  HCl, and 3% THB with 9 mol L<sup>-1</sup> HCl was utilized for As(III) and total As, respectively. The system utilized an integrated reactor and gas–liquid separator, allowing improved sensitivity and interference free determination.

Selective retention of As(V) by an anion-exchange resin and total As after  $KMnO_4$ oxidation was also proposed [47]. Sequential combination of two different columns, in which, the first selectively retains As(III)–pyrrolidinedithiocarbamate (PDC) complex, while the second retains As(V), was reported for ICP–AES without HG [162]. Simultaneous determination of As(III) and As(V), based on the reduction kinetics using a co-centric HG, was also reported [163].

#### 4.5 HG–GFAAS techniques

An alternative approach to the use of a heated quartz tube atomizer is the deposition and collection of the volatile hydrides inside a preheated graphite furnace. The first report on the use of GF atomizer for the atomization of hydrides was by Knudson and Christian [164] in 1974. Since then, HG coupled with GF atomization technique has been carried out by several workers, and it has been reviewed [165]. Kalahne *et al.* [166] reported a comparative study of the performances of HG coupled to GFAAS, QFAAS, and ICP–AES. HG–GFAAS showed the lowest  $LOD$  (3 ng L<sup>-1</sup>); however, HG–GFAAS suffered severe interference from other hydride-forming elements than the other two detectors. There are two approaches to using graphite furnace atomization; online atomization and in situ trapping of hydrides in the furnace. The in situ trapping technique has been shown to enhance the sensitivity significantly and eliminate efficiently the oxidation-state influence on the signal [167]. A comparison of both the techniques was reported [168].

A comprehensive review on FI techniques coupled to GFAAS was presented by Fang and Tao [169]. Various approaches of elemental speciation using GFAAS were also reviewed [170].

The first report on speciation analysis using HG–GFAAS was by Shaikh and Tallman [171]. They determined  $As(III)$ ,  $As(V)$ , MMA, and DMA utilizing a U trap coupled to GFAAS. Other reported methods on arsenic speciation analysis were based on selective HG, solvent extraction and/or sequential volatilization utilizing the cold trap. Selective HG from As(III) species and total As after reduction was reported by several authors [120–122, 124, 172]. SPE using APDC [173] and 2,3-dimercaptopropane-1-sulfonate on  $C_{18}$  cartridge was reported [174]. Co-precipitation of As(III) with Cu–PDC and total arsenic after L-cysteine reduction was reported with a  $ng L^{-1}$ LOD [175]. Russeva *et al.* [176] separated As(V) selectively on a column packed with organotin material. Barrera *et al.* [177] reported a selective HG method to determine the four arsenic species. Wille [178] reported a method to distinguish between hydride active and inactive species, i.e. first-order speciation, based on online UV oxidation/ microwave digestion in alkaline  $K_2S_2O_8$  media. This showed that a significant fraction of As in seawater contains non-hydride active species. Sturgeon et al. [179] determined inorganic, methylated, and organoarsenicals such as arsenobetaine (AB) and arsenocholine (AC) based on selective reaction media and ion exchange with a UV irradiation procedure in river-water reference materials. They reported [179] that 22% of total arsenic was present in the non-hydride active form. Cabon and Cabon [54] reported an FI–HG procedure based on selective reaction media and online UV treatment in alkaline  $K_2S_2O_8$  media to determine hydride active and inactive fractions in seawater. About 15% of total arsenic was in the non-hydride active form. An LOD of  $1.5$  ng L<sup>-1</sup> was reported, with a pre-concentration factor of 1000. Hermelo *et al.* [180] reported the selective online formation of As(III)–PDC complex in a knotted reactor and GFAAS detection with an LOD of  $8 \text{ ng L}^{-1}$ .

The role of modifiers to improve the performance of GF was investigated by various authors. The sensitivity of various As species were reported to differ according to the volatility of hydrides, i.e. less volatile hydrides gave a more intense signal than that of highly volatile hydrides. The addition of Ni was reported to eliminate this effect [181]. Slaveykova et al. [182] investigated the atomization behaviour of arsenic species in pyro-coated and tungsten-coated graphite tubes with Pd modifiers and reported that it improved the sensitivity of various As species. Barrera et al. [177] investigated the hydride-trapping efficiency with Ir-, W-, and Zr-coated graphite tubes, and reported that Zr-coated tubes gave a better performance. A comparative study of matrix modifiers in GFAAS was reported by Niedielski et al. [183].

## 4.6 HG–AFS techniques

Tsuji and Kugu [184, 185] first reported HG coupled with AFS. Mester and Foder [186] studied the characteristics of AF signals of As species and emphasized the need for independent calibration for each As species. D'Ulivo et al. [187] discussed the methods to improve the performance of HG–AFS techniques using additives and optical fibres. AFS detectors are very sensitive and selective for hydride-forming elements than AAS and AES. An overview of the speciation analysis of As by AFS was reported by Cai [17].

El-Hadri et al. [55] determined As(III) with 3% THB, and 2 mol  $L^{-1}$  of HCl and total As after prereduction with KI and species concentrations were calculated from the slope of the calibration curves. Lee et al. [188] described a method based on cation–anionexchange columns connected in series. Lopez and Castro [189] utilized FI coupled with pervaporation prior to HG for dirty water samples containing highly suspended matter. van Elteren et al. [53] reported a procedure based on the co-precipitation of As(III) dibenzyldithiocarbamate. Yan et al. [50] reported a FI–HGAFS technique, in which, As(III) was selectively complexed with APDC online, and total As was determined following L-cysteine prereduction. Castro et al. [190] reported a method to estimate total hydride active As species and total As including 'hidden As' after UV irradiation. AB was mineralized only in alkaline  $pH > 11$  with an irradiation time of 60 min. Gong et al. [44] reported an SPE method for As(V) selective extraction. Featherstone et al. [191] reported a HG–CT–AFS method for the determination of four As species with a ng L<sup>-1</sup> level LODs. Leal et al. [192] proposed a new technique based on multisyringe

flow injection (MSFI), in which As(III) and total arsenic were determined simultaneously after KI–ascorbic acid reduction.

#### 4.7 Electrochemical hydride generation (EcHG) techniques

Electrochemical hydride generation, as a sample-introduction system to atomic spectrometry, is an alternative to wet chemical HG. Several advantages have been reported over wet chemical methods and the topic being reviewed [193, 194]. It has widely being used for total element determination, but for speciation analysis little information is available. The efficiency of EcHG depends mainly upon the cathode material, electrolytic current, dimensions of the flow-through cell, and gas–liquid separator. Cathode materials like Pt, Au, Ag, and glassy carbon generates hydride only from the lower valency state of the hydride-forming elements, whereas Cd, Hg–As, and Pb generate hydrides from a higher valency as well, with different efficiencies [194].

Ding and Sturgeon [195] reported that the efficiency of arsine formation varied according to the nature of the cathode material and reaction matrix. Thus, As(V) could be reduced to AsH<sub>3</sub> using lead as cathode in  $HCl + H<sub>2</sub>SO<sub>4</sub>$  media without prereduction. Further, the addition of a high concentration of hydroxylamine hydrochloride completely suppressed the signal of As(V), but MMA and DMA were reported to interfere. Schaumloff and Neidhart [196] selectively determined As(III) using a fibrous carbon cathode and total As after online reduction with L-cysteine. Pyell et al. [197] reported the use of fibrous carbon and lead as cathode materials to determine As(III) and total arsenic, and employed a cold trap to remove the interference due to methylated species. Li et al. [198] reported a procedure for inorganic As speciation analysis, based on the HG efficiency at 0.6 and 1.0 A electrolytic currents using a glassy carbon cathode. The concentrations of species were calculated from the slope of the calibration curves.

#### 4.8 Speciation analysis using microorganisms

The ability of microorganisms to incorporate elemental species (biosorption) became a useful technique in speciation analysis. The literature on biological substrate for metal preconcentration was reviewed by Madrid and Camara [199] and Zylkiewicz [200].

Calzada et al. [25] reported that As(V) was biosorbed by the alga Chlorella vulgaris, whereas other species were unadsorbed. Smichowski et al. [201] successfully utilized Saccharomyces cerevisine (baker's yeast) for the selective retention of As(III) in a batch system and quantified by HG–ICP–AES. Contrary to this, Koh *et al.* [202] reported that As(V) was retained better than As(III) in an immobilized column. They reported that in an immobilized column, the metal-binding (–SH) groups were blocked, and so As(III) was not adsorbed.

#### 5. Selenium speciation analysis

Selenium as an essential trace element has attracted increasing attention in recent years, as evidence for its involvement in human health has become apparent. Selenium exists in the environment in several oxidation states as well as a variety of inorganic and organic compounds. Dissolved inorganic selenium can be found in natural waters as selenide ( $-I$ I), as colloidal elemental Se(0), as selenite ion Se(IV), and as selenate anion Se(VI). Organic forms of Se that may be found in the aqueous environment are volatile (methyl selenides) or non-volatile (trimethyl selenonium ion, seleno-amino acids, and their derivatives) [203].

As only Se(IV) is reduced to SeH<sub>2</sub> with THB, the other species should be converted into  $\mathcal{S}(IV)$  in order to generate  $\mathcal{S}(H_2)$ , by performing an analysis using three separate aliquots, i.e. (a) with no further treatment—direct determination of  $Se(IV)$ , (b) after reduction of Se(VI) to Se(IV) with HCl or  $HBr/KBr-Br<sub>2</sub>$ —the sum of Se(IV) and Se(VI), Se(VI) thus being determined by difference, (c) mineralization of organic matrix by UV-irradiation or wet digestion followed by reduction to Se(IV)—determination of total Se. The difference between the total Se and the sum of  $Se(V)$  and  $Se(V)$  is attributed to organic selenium compounds. However, it has been reported recently that the presence of dimethylselenium (DMeSe) and dimethyldiselenium (DMeDSe) compounds also contributes to the Se signal in HG methods [204].

## 5.1 HG-AAS and ICP-AES techniques

Many workers have determined inorganic selenium species, i.e. Se(IV) and Se(VI), utilizing the formation of  $\text{SeH}_2$  only from the  $+IV$  state. Se(VI) is reduced to the +IV state by heating/boiling with  $4-6$  mol L<sup>-1</sup> of HCl [205–209] or with bromide ions [210, 211].

Cutter [79, 212] introduced a 'U' cold trap at liquid-nitrogen temperature to concentrate the evolved SeH<sub>2</sub> before atomization and achieved ng  $L^{-1}$ -level LODs. Yu *et al.* [85] proposed selective retention of  $Se(IV)$  on thiol cotton, and  $Se(VI)$  was determined after reduction with HCl and TiCl<sub>3</sub>. Flores *et al.* [213] determined Se(IV) using a simple microbatch venturi type system, which was reported to consume less of the reagents. Maleki et al. [214] proposed a procedure in which the sample was injected into the reactor containing solid THB and tartaric acid. Se(VI) was determined after prereduction with L-cysteine. This method utilizes only solid reagents, thus obviating reagent preparation, and has an extended linear working range (up to  $1200 \mu g L^{-1}$ ). Brunori *et al.* [78] reported a microwave prereduction method for Se(VI) coupled with CF–HGAAS. Selenocysteine and selenomethionine were partially reduced to Se(IV); hence their removal by cation exchange resin was recommended for Se speciation analysis.

Narasaki and Mayumi [70] extracted Se(IV) selectively with 4-nitro-o-phenylenediamine into toluene as 4-nitropiazselenol. The extract was mineralized with  $HNO<sub>3</sub>$ – HClO4 and subjected to HG–ICP–AES detection. Total Se was determined after HCl reduction. Stripeikis et al. [215] reported a method to remove Fe and Cu interferences by retention on Dowex IX-8 resin as their chloro complexes, whereas Se species were not retained. The eluted Se species were passed through a heated PTFE coil for online reduction and determined by HG–ICP–AES. Schermer et al. [216] selectively determined Se(IV) by careful optimization of sample flow rate and electrolysis current in EcHG with microwave-induced plasma AES detection. In addition, the selective determination by complexation and solvent extraction has also been reported. Morabito [10] reviewed extraction procedures applied to speciation analysis.

Reaction of aromatic o-diamines with Se(IV) to form the corresponding selenodiazoles (piazselenols) is one of the most commonly used derivatization procedures. As only Se(IV) forms the piazselenol derivative, Se(VI) could be determined as the difference between total Se and Se(IV) similar to HG techniques. Tamari and Ogura [217] determined Se(IV) selectively by HGAAS, after extracting the complex Se(IV)- DAN (2,3 diaminonapthalene) into cyclohexane, decomposing the organic phase by  $HNO<sub>3</sub>-HCCO<sub>4</sub>$  acids, and then reducing all Se to Se(IV) with HCL. Se(IV) with DAN (2,3-diaminonapthalene), extracted the complex with cyclohexane and determined by HGAAS, after destructing the organic phase with  $HNO<sub>3</sub>-HClO<sub>4</sub>$ . Total Se was determined after HCl prereduction. This procedure allowed an LOD at ppt levels. Reddy et al. [77] reported a method for the selective co-precipitation of Se(IV) on CuO at pH 5.5 and desorbed at pH 12.5. Total Se was determined after HCl prereduction.

## 5.2 FI–HGAAS techniques

The introduction of FI systems offers efficient online sample prereduction methods, utilizing chemical, microwave, and photochemical methods. This avoids drawbacks such as analyte loss due to volatilization, reduction to elemental state, etc. associated with conventional offline methods. A detailed discussion of microwave digestion procedures for environmental samples has been published [218].

Fernandez et al. [73] determined Se(IV) using 0.5% THB and  $4 \text{ mol L}^{-1}$  HCl, and total Se was determined using  $6 \text{ mol L}^{-1}$  HCl and heating the sample to 140°C to convert all  $Se(VI)$  to  $Se(IV)$  online. Online preconcentration of  $Se(IV)$  by co-precipitation with La(OH)<sub>3</sub> was reported with an LOD of  $1 \text{ ng } L^{-1}$  [219]. Rubio et al. [220] described an online UV-irradiation technique for reduction of Se(VI). Online reduction of Se(VI) utilizing the microwave-irradiation technique has been reported by several authors [221–224]. Brunori et al. [78] reported a microwave-reduction method for Se(VI) coupled with CF–HGAAS. Seleocysteine and selenomethionine were partially reduced to  $Se(IV)$ ; hence their removal using a cation-exchange resin was recommended prior to HG. Burguera et al. [225] utilized a time-based injector for injecting  $4 \text{ mol L}^{-1}$  HCl for Se(IV) and microwave irradiation in  $12 \text{ mol L}^{-1}$  HCl for total Se determination. They reported that it avoids the problems associated with conventional microwave irradiation. LaFuente et al. [226] reported an online focused microwave digestion method for the determination of Se(IV), Se(VI), and organic Se. Organic Se was decomposed first by  $HBr/KBrO<sub>3</sub>$  reagent with microwave irradiation for total Se determination. Gallignani et al. [227] reported an online sequential MW irradiation method in which a 10% v/v mixture of HCl and HBr medium was used. Mendez et al. [228] reported a time-based UV and ultrasound (US) irradiation procedure for the reduction of  $Se(VI)$  and organic Se using  $KNO<sub>3</sub>$  as a photoinhibitor. In spite of the claimed advantages, low recoveries of Se were reported from samples contain high organic matter [229]. Stripeikis et al. [230] used a conventional water bath for online reduction of Se(VI) using  $6.8 \text{ mol L}^{-1}$  HCl and reported that it was advantageous over microwave heating.

Selective retention of inorganic Se species prior to the HG step has also been reported. Pyrzynska [231] selectively preconcentrated Se(IV) and Se(VI) on cellex-T anion-exchange resin. Se(IV) was selectively eluted with  $0.01 \text{ mol L}^{-1}$  HNO<sub>3</sub>, whereas Se(VI) was eluted with  $4 \text{ mol} L^{-1}$  HNO<sub>3</sub>. Selective retention on activated alumina

and successive elution with  $1 \text{ mol L}^{-1} \text{ NH}_3$  and  $4 \text{ mol L}^{-1} \text{ NH}_3$  was also reported [232]. Carrero and Tyson [233] immobilized THB and Se(IV) selectively on an anion-exchange resin,  $\text{SeH}_2$  was generated by injecting HCl with QFAAS detection. The immobilization of THB improved the  $\text{SeH}_2$  generation efficiency and reduced the blank signal. An LOD of  $0.12 \mu g L^{-1}$  was achieved with a preconcentration time of 3 min. Itoh *et al.* [74] reported selective retention of  $Se(IV)$  on an anion-exchange resin loaded with bismuthiol-II. Larraya et al. [76] reported an online selective retention of Se(IV) on an alumina-filled microcolumn and total Se after reduction with HCl online. Se(IV) was eluted with  $2 \text{ mol} L^{-1}$  NH<sub>3</sub>. The reported LOD was  $6$ ng L<sup>-1</sup>. Ornemark and Olin [75] reported a preconcentration procedure using Dowex IX8, in which both Se(IV) and Se(VI) were retained. Se(IV) was retained when the sample pH was  $> 8$ , and Se(VI) was retained at a pH between 1 and 10 with 1 and  $5 \text{ mol L}^{-1}$  HCl used for elution. Selective retention of Se species on XAD-8 column was also reported [234]. Gomez-Ariza et al. [235] reported the simultaneous retention of  $Se(IV)$ ,  $Se(VI)$  and  $Se(-II)$  on an XAX anion-exchange resin and eluted with  $1 \text{ mol L}^{-1}$  of HCOOH, HCl, and CS<sub>2</sub>, respectively. Sahin et al. [236] reported the selective retention of Se(IV) on mercaptosilica and eluted with  $0.2\%$  KIO<sub>3</sub> and 1 mol L<sup>-1</sup> HCl.

#### 5.3 HG–ETAAS techniques

Rodden and Tallman [237] reported Se speciation analysis using HG–CT–GFAAS detection, in which the sample was passed through an XAD-8 anion exchange resin at pH 1.6–1.8 for organic-matter removal. Total Se was determined after HCl reduction. Cabon and Erler [71] utilized UV irradiation in basic medium and HCl to reduce  $Se(-II)$  and  $Se(VI)$  to  $Se(IV)$ , and attained LODs of 10, 21, and  $13 \text{ ng } L^{-1}$  for Se(IV), Se(VI), and Se(-II), respectively, with a preconcentration factor of 1000. Li and Deng [238] reported a procedure based on the selective adsorption of Se species on TiO<sub>2</sub>, and subsequent determination by GFAAS. Saygi et al. [239] determined Se(IV) after the selective separation of the Se(IV)–PDC complex on Diaion HP-2MG and total selenium after HCl reduction. Chung et al. [120] reported the selective extraction of  $Se(IV)$  with APDC in  $CCl<sub>4</sub>-CHCl<sub>3</sub>$ and Se(VI) after TiCl<sub>3</sub>–HCl reduction. Selective extraction of Se(IV) using APDC–MIBK was also reported [124].

Stripeikis *et al.* [72] reported an online separation and preconcentration procedure, in which Se species were preconcentrated on Dowex IX8 anionic resin, which, on elution with 0.1 and  $4M$  HCl, separated Se(IV) and Se(VI) species. Interference due to chloride ions during atomization was reduced using iridium-coated graphite tubes. An LOD of  $10 \text{ ng } L^{-1}$  was reported. Niedzielski *et al.* [84, 240] reported the determination of Se(IV) directly and total selenium after HCl reduction with palladium modifier. Sahin et al. [241] studied the effect of Ni and  $Pd + Mg$  modifiers in Se speciation analysis and proposed that the presence of  $HNO<sub>3</sub>$  is necessary to obtain equal sensitivities from Se(IV), Se(VI), selenocysteine, and selenomethionine. Fragueiro et al. [242] reported a novel extraction method based on single-drop microextraction (SDME) of  $\text{Set}^1_2$  generated in a headspace, which has subsequently been injected into graphite furnace AAS. Total Se was determined after

UV irradiation. Tuzen et al. [243] reported a method based on the selective co-precipitation of Se(IV) on  $Mg(OH)_{2}$ .

## 5.4 HG–AFS techniques

As AFS techniques are very sensitive, they can be directly used without a preconcentration procedure. HG-AFS techniques reported for Se speciation analysis are also based on the formation of  $\text{SeH}_2$  only from the  $+$ IV state, and  $\text{Se(VI)}$  is calculated as the difference from total Se. He et al. [244] reported a direct determination procedure by CF in which higher flow rates  $(22 \text{ mL min}^{-1})$  of sample were used to increase the sensitivity. Se(IV) was determined with  $1.5 \text{ mol L}^{-1}$  HCl and  $1.5\%$  of THB, whereas the total Se was determined after reduction with  $4\%$  KBr and  $1.5 \text{ mol L}^{-1}$ . An LOD of 0.96–1.3 ng L<sup>-1</sup> was reported. Eva Moreno *et al.* [67] determined Se(IV) directly and total Se after HCl reduction by FI–HGAFS. Lu et al. [245] reported a preconcentration procedure in which the Se(IV)–PDC complex is formed online and adsorbed on a polyterafluoroethlene (PTFE) fibre-packed micro-column. THB is used as eluent, and its subsequent reaction with HCl generates SeH<sub>2</sub>. An LOD of  $4 \text{ ng L}^{-1}$ was reported. Bryce *et al.* [222] reported an online focused microwave reduction method to determine Se(VI), whereas Se(IV) was determined directly. These authors reported another method based on SPE, in which both Se(IV) and Se(VI) were retained and sequentially eluted with formic acid and HCl.

Chen *et al.* [246] investigated the photochemical behaviour of  $Se(V)$ ,  $Se(VI)$ , and organic selenium compounds by UV irradiation. Se(IV) was determined with  $3 \text{ mol L}^{-1}$ HCl, and organic Se was determined in the matrix of  $1\%$  HNO<sub>3</sub> and  $2\%$  HCl with 2.5 h of UV irradiation at 300 nm. Total Se was determined after refluxing the sample from UV irradiation. Mendez et al. [228] reported a time-based combination of UV and ultrasound irradiation techniques for Se(VI), selenocysteine, and selenomethionine.  $KNO<sub>3</sub>$  was used as photoinhibitor. Tang *et al.* [69] reported the preconcentration of  $Se(IV)$  using co-precipitation with  $La(OH)_{3}$ .  $Se(IV)$  is redissolved with HCl, and total Se was determined after HCl reduction. An LOD of  $3 \text{ ng } L^{-1}$  was reported.

#### 5.5 Speciation analysis using microorganisms

The use of common yeast cells (Saccharomyces cerevisiae) for selective retention of  $Se(IV)$  was reported by Perez-Corona *et al.* [247], whereas  $Se(VI)$  was not biosorbed. Robles et al. [248] utilized living bacteria (Pseudomonas putida and Escherichia coli) for the separation of Se(IV) and Se(VI) species.

#### 5.6 Studies on the prereduction of  $\mathcal{S}e(VI)$  to  $\mathcal{S}e(IV)$

Determination of inorganic Se species by HG methods requires the analyte to be present in the  $+IV$  state; hence Se(VI) could be determined differentially if total Se is determined after prereduction. It is generally accomplished by heating the analyte in the presence of HCl usually at an acid strength of  $4-6 \text{ mol L}^{-1}$ . The reaction is as follows:

$$
HSeO_4^- + 3H^+ + 2Cl^- \leftrightarrow H_2SeO_3 + Cl_2(aq) + H_2O.
$$
 (1)

However, there are very conflicting statements in the literature as to the appropriate conditions for the reduction of Se(VI) with HCl. The controversies concern the optimal acid concentration, reaction time, and temperature. Pivonka et al. [249] presented a table which clearly demonstrated the differences in the recommended conditions. There are probably several reasons for the controversies. The most likely reason is that the sample solutions have contained other reagents possessing redox properties and that the presence of these will alter the redox potential of the HCl solution, demanding different concentrations of HCl, heating periods, and temperatures.

Cutter [79, 212] recommended that the reduction be carried out in boiling  $4 \text{ mol L}^{-1}$ HCl for 15 min and warned that prolonged boiling could lead to a reduction to an elemental state. However, the conclusion was challenged on thermodynamic grounds by Bye [250], who suggested that the loss of Se from samples, which were boiled for too long, might be due to the formation of volatile  $\text{Se}(IV)$  compounds such as  $\text{SeCl}_4$  or SeOCl<sub>2</sub>. But, Krivan *et al.* [251], in a study using radiotracers, showed that the loss of Se was due to the back-oxidation by  $Cl<sub>2</sub>$ , which in turn had formed from the reaction of HCl with strong oxidizing agents like  $H_2O_2$ . Bye [250] also concluded that the important component of the reducing solution was the  $Cl^-$  ion and that the reduction could be carried out quantitatively in con. HCl solution. Brimmer et al. [252] investigated the optimal reduction conditions using HCl and concluded that heating at 91 $\degree$ C in 6 mol L<sup>-1</sup> HCl for 30 min resulted 99.9% reduction of Se(VI). The authors showed that the reaction is first-order  $(k=5.8 \times 10^{-4}$  and  $3.9 \times 10^{-3}$  s<sup>-1</sup> for 4 and  $6 \text{ mol L}^{-1}$  HCl) with respect to Se(VI), and the reaction rate increased sharply with HCl concentration. No loss of Se was observed when closed vessels were used, which supports the possible formation of volatile chloride species. Contrarily, Hill *et al.* [253] reported that reduction should preferably be carried out in open vessels, to facilitate the escape of Cl<sub>2</sub> generated, which otherwise may result in back-oxidation. Bye and Lund [254] reported that complete reduction of Se(VI) was achieved in 10 mol  $L^{-1}$  HCl heated to 60°C for 15 min, and so boiling is not necessary. According to Bye and Lund [254], the activation energy,  $E_A$ , was 83 kJ mol<sup>-1</sup>, and the pseudo-first-order rate constant of the reaction was  $6.2 \times 10^{-5}$  and  $3.3 \times 10^{-4}$  s<sup>-1</sup> for 60 and 80°C using 4 mol L<sup>-1</sup>. This is in good agreement with the values reported by Brimmer et al. [252]. The standard enthalpy change ( $\Delta H_f$ ) was found to be 116 kJ mol<sup>-1</sup>, thus indicating an endothermic reaction, and so when the temperature is increased, the equilibrium constant will also be increased, leading to more efficient reduction.

Petterson and Olin [255] reported that a 99.9% reduction in 30 min required heating to 105, 85, and 65 $^{\circ}$ C for 4, 5, and 6 mol L<sup>-1</sup> HCl. Kinetic calculations showed that  $E_A$  was  $126 \pm 10 \text{ kJ} \text{ mol}^{-1}$ , about  $40 \text{ kJ} \text{ mol}^{-1}$  higher than that of Bye and Lund [254]. Furthermore, the authors pointed out that the reaction order was almost 5 at a constant chloride concentration  $(4-5 M)$  and a constant ionic strength of 5 M, and is greater than the stoichometric coefficient for  $H^+$  in reaction (1). According to Petterson and Olin [255], the high dependence of the reaction rate on acidity indicates that some kind of selenate-proton cluster might be formed before the selenium atom is attacked by chloride. Hill et al. [253] reported that the activation energy  $E_A$  was 90.4 kJ mol<sup>-1</sup>, and complete reduction of Se(VI) to Se(IV) could be achieved by heating to  $70^{\circ}$ C for 6 min using 6 mol  $L^{-1}$  HCl.

The prereduction has also been accomplished by heating with bromide ions, and it has been reported to offer several advantages over chloride ions. It prevents vigorous reactions, but shows better interference control [256] and improvement in flow

characteristics in a CF system, resulting in a better stability and reproducibility of AFS signals [244]. It also has an improved sensitivity in CF–HGAAS, as reported recently [257]. He *et al.* [244] reported that solid NaBr could be added to the samples, and so dilution caused by HCl could be avoided.

The equilibrium constant of reaction (1), when  $Br^-$  is used instead of Cl<sup>-</sup>, is 1.1 kg<sup>4</sup> mol<sup>-4</sup>, whereas with Cl<sup>-</sup>, it is  $7.1 \times 10^{-11}$  kg<sup>4</sup> mol<sup>-4</sup> [258]. This suggests that the prereduction reaction with HBr is more favourable than with HCl, and, the back-oxidation caused by  $Cl_2$  could be avoided. For example, D'Ulivo *et al.* [256, 259] reported that organic selenium compounds when digested with  $HBr/Br<sub>2</sub>$  converted all Se to Se(IV) only. Despite these advantages, reports on the prereduction using  $Br^-$  are fragmentary. Brindle and Lugowska [258] reported that bromide ions are found to be 19 times more efficient than chloride ions. They pointed out the particular reduction power of halide ions toward Se(IV), overcoming the kinetic inhibition of the prereduction of  $Se(VI)$ , may be connected with the mechanism of reaction (1):

$$
SeO_4^{2-} + 8Cl^- + 8H^+ \leftrightarrow SeCl_6^{2-} + Cl_2 + 4H_2O.
$$

Se(IV) halides may be hydrolysed in aqueous solutions to  $H_2$ SeO<sub>3</sub>.

In all these studies reported so far, the role of halide ions was assumed to be that of a reducing agent. Kenduzler et al. [260], in a recent kinetic study, proposed that the initial step of the conversion of  $Se(V)$  to  $Se(V)$  in the presence of  $Br^-$  is a nucleophilic substitution of the protonated –OH group of selenic acid in an acid-catalysed reaction. According to the authors, such a nucleophilic substitution reaction would explain the smaller reactivity of  $Cl^-$  ions compared with  $Br^-$  ions.

#### 6. Antimony speciation analysis

Antimony is a cumulative toxic element that has been detected in natural waters at ultra-trace levels [261]. In natural waters, it may exist as  $Sb(III)$  or  $Sb(V)$ , and a variety of methylated forms. In seawater at pH 8, both oxidation states are strongly hydrolysed, leading to the formation of  $Sb(OH)_{3}$  and  $Sb(OH)_{6}^{-}$ . These two inorganic forms exhibit pronounced differences in their analytical behaviour, toxicity, and mobility [34]. Trivalent species are reported to be more toxic than pentavalent forms. In seawater, methylated species represent about 10% of the total dissolved Sb, with the monomethyl species being predominant [34].

The determination of antimony species in environmental water samples often requires a preconcentration step, because of its presence at  $ng L^{-1}$  levels. Hence, various preconcentration methods utilizing liquid–liquid extraction (LLE), solid-phase extraction (SPE), and coprecipitation methods precede atomic spectrometric detectors. Smichowski et al. [33] published a review on the techniques of antimony speciation in 1998, and Nash *et al.* [34] presented an extensive review on the methodologies for antimony determination in environmental samples in 2000. Both the reviews include various preconcentration methods combined with atomic spectrometric detectors for speciation analysis. Hence, we review the procedures reported after the year 2000.

## 6.1 Speciation analysis based on selective HG

Methods based on selective hydride formation are attractive, as there are no additional steps involved. This method relies on the pH dependency of stibine formation from Sb(III) and Sb(V) species. Sb(III) forms  $SbH_3$  over a wide range of pHs, i.e. 2–8, and total Sb is determined under strongly acidic conditions ( $\approx$ 4 mol L<sup>-1</sup> HCl) or with prereduction. Nakashima et al. [262] utilized this procedure first and reported the selective determination of Sb(III) in 0.35 mol  $L^{-1}$  HCl and 4 mg mL<sup>-1</sup> of  $Zr(IV)$ , with ng  $L^{-1}$  LODs. This pH-dependent reduction of THB has been utilized by many authors [86, 101, 103, 111, 263] to selectively determine Sb(III) and total Sb after prereduction. Apte and Howard [264] utilized the cryogenic-trap method to concentrate the hydrides and attained 1 and  $10 \text{ ng } L^{-1}$  LODs for Sb(III) and total Sb. Andreae et al. [265] reported a trap method for methylated species which differ in their boiling points. Various buffers and acids were reported to the selective determination of Sb(III), e.g. citric acid (pH 2) [101, 103], acetate buffer [266], and phosphoric acid (pH 1.81) [267].

Nakahara and Kikui [268] reported a sequential determination of Sb(III) and total Sb in maleic and tartaric acid with an LOD of  $0.19 \mu g L^{-1}$ . Selective determination of Sb(III) in 0.1 mol  $L^{-1}$  (pH 5.5) or 6% citric acid and total Sb after KI reduction was also reported [269, 270]. Garcia et al. [271] reported a continuous tandem online separation of Sb(III) as its PDC complex and extraction into methylisobutyl ketone (MIBK) phase and aspiration into ICP. Sb(V) was determined after KI reduction of the aqueous phase. However, the reported LOD of  $3 \mu g L^{-1}$  was not sufficient for real samples without preconcentration. Feng et al.  $[272]$  reported a method to determine Sb species based on the reduction kinetics with L-cysteine. Ulrich *et al.* [273] determined Sb(III), Sb(V) and trimethylstiboxide (TMeSbO) using fluoride as matrix modifier with LOD of 1.1, 1.2, and  $1.4 \mu g L^{-1}$ , respectively.

Selective extraction of Sb(III) species using APDC–MIBK [121, 124, 274] and lactic acid–Malachite green [270, 275] have been reported. Similarly, SPE using a variety of adsorbents has been reported. Yu et al. [85] reported the selective retention of Sb(III) on thiol cotton, with HGAAS detection. Sb(V) was reduced with KI–ascorbic acid, and an LOD of  $5 \text{ ng } L^{-1}$  was reported. Thionalide loaded on a silica-gel column for selective retention of Sb(III) was also reported [276].

## 6.2 FI–HGAAS techniques

FIHG techniques utilizing online preconcentration and/or selective reduction procedures have been applied to Sb speciation analysis. Guntinas et al. [270, 277] used 6% citric acid for Sb(III) and  $4 \text{ mol L}^{-1}$  HCl media to determine total Sb and reported an LOD of  $7 \text{ ng } L^{-1}$ . Garbos *et al.* [82] reported the use of  $0.02 \text{ mol } L^{-1}$  HCl for Sb(III). Cabon and Medac [81] used a 'U' trap filled with chromatographic phase for concentration and used Tris-HCl buffer (pH 7.2) for Sb(III) determination with LOD of  $20 \text{ ng } L^{-1}$ . Selective sorption of Sb(III) on Duolite GT-73 with -SH functional groups coupled to segmented FI–HGAAS was reported by Erdem and Erogulu [278] with an LOD of  $60 \text{ ng L}^{-1}$ . Zheng *et al.* [279] used a nanometer-sized TiO<sub>2</sub> to preconcentrate Sb(III) and Sb(V) and attained an LOD of  $50 \text{ ng L}^{-1}$ .

## 6.3 HG–GFAAS and AFS techniques

HG *in situ* preconcentration on graphite tubes offers another efficient tool for antimony speciation analysis. A comprehensive review of this subject was reported [165]. Smichowski et al. [280] reported a method for the simultaneous preconcentration of Sb(III) and Sb(V) based on the selective retention on an alumina microcolumn under controlled pH conditions using phosphoric acid. Garbos *et al.* [82] reported a pH-based selective adsorption procedure for Sb(III) and total Sb using polyorgs 31 with amidoxime and amino functional groups, with an LOD of 30 ng  $L^{-1}$ . Selective stibine generation from  $\text{Sb}^{3+}$  and *in situ* trapping on graphite tubes were reported by several authors [84, 86, 172] with  $ng L^{-1}$  LODs. Garbos *et al.* [82] reported a selective preconcentration of Sb(III) as PDC complex, sorbed over  $C_{16}$  bonded silica jel, with an LOD of  $7 \text{ ng } L^{-1}$ . Chung et al. [120] reported the selective extraction of Sb(III) as its PDC complex in  $\text{CCl}_4$ –CHCl<sub>3</sub> and Sb(V) with TiCl<sub>3</sub>–HCl reduction. Cabon and Madec [81] utilized the selective determination of Sb(III) using  $0.2 \text{ mol L}^{-1}$  Tris-HCl buffer and total Sb after online UV oxidation, and reported that oxidation led to a better recovery than reduction.

AFS has been demonstrated as a sensitive technique for hydride-forming elements having its resonance lines below 250 nm [186]. Determination of Sb by HGAFS has been described in detail by D'Ulivo et al. [281]. Tsujii [282] reported the selective determination of Sb(III) using HF as a masking agent for Sb(V). Selective determination of Sb(III) with acetate buffer (pH 5) was reported by Suo and Huang [283]. Deng *et al.* [83] used 8-hydroxyl quinoline as a masking agent for  $Sb(V)$  in the determination of Sb(III). Sb(V) was determined after KI prereduction, and total Sb was determined after microwave digestion and KI prereduction. The difference between total Sb and Sb(III) and Sb(V) was attributed to organic Sb. El-Hadri et al. [55] reported a method to determine Sb(III) and Sb(V) based on the efficiency of hydride generation with and without KI reduction.

#### 7. Tellurium speciation analysis

Tellurium is regarded as a rare, toxic, non-essential element. In environmental water samples, it occurs normally at ng  $L^{-1}$  levels. Tellurium can exist as Te(IV), Te(VI), and, very rarely, dimethyl telluride and dimethyl ditelluride [39]. Studies on its speciation in natural waters are scarce. Only inorganic speciation in some seawater samples has been reported. A detailed review on its determination in environmental samples has been reported [39].

As tellurium occurs at  $ng L^{-1}$  levels, its determination invariably requires a preconcentration step. APDC, sodium diethyldithiocarbamate, and dithiozone are the ligands usually employed in LLE prior to detection [284]. The solvent extraction of Te(IV) has been reviewed [285].

Andreae [286] reported a method in which both Te(IV) and Te(VI) were coprecipitated with  $Mg(OH)_2$ , which was then redissolved in HCl and determined by HG–GFAAS. Total Te was determined after reduction with HCl. Lee and Edmond [287] used alkaline co-precipitation for preconcentration of Te(IV) and determined by HG–GFAAS with in situ trapping. Yu et al. [85] reported the selective sorption of  $T_{e}(IV)$  on thiol cotton with HGAAS detection. Chung *et al.* [120] reported the selective extraction of Te(IV) as PDC complex into  $CHCl<sub>3</sub>-CCl<sub>4</sub>$  with GFAAS detection. Te(VI) was reduced with  $TiCl<sub>3</sub>-HCl$  and determined as  $Te(IV)$ . Yoon *et al.* [288] determined Te species by *in situ* trapping of TeH<sub>2</sub> in graphite tubes. Total Te was determined after HCl reduction. Korez et al. [289] reported the selective retention of  $Te(V)$  on mercaptosilica and total Te determination after HCl reduction.

## 8. Trends and future prospects

Hydride generation is the oldest method used for vaporization of trace elements. HG combined with atomic spectrometric detection has become the established method for As, Sb, Se, Te, etc. The technique comprises several distinct stages, notably generation of the volatile analyte, its collection (optional) and transfer to the atomizer, and atomization. During the last three decades, significant developments have taken place, which have improved the overall efficiency of the technique. Thus, ppb/sub-ppb levels of elements such as As and Se could be determined without the use of preconcentration (pre- or post-HG) techniques. However, a preconcentration step is necessary for elements such as Sb and Te, as their levels in the environment are normally at ppt levels. Hence, various preconcentration techniques coupled with HG remain an active area of research. Furthermore, an enhancement of the signal-to-noise ratio can be secured by collecting the species in a cryogenic trap prior to its introduction to the detector, thereby minimizing any dilution effects induced from the presence of co-evolved gases and other gases used to achieve phase separation and transport. One of the main advantages of cryogenic trapping technique is that the species do not come into contact with any liquid or solid sorbent materials, thus significantly reducing the possibility of their alteration/decomposition, but its bulkiness and the need to handle cryogenic liquid pose major drawbacks. HG coupled with in situ traping in GF atomizer offers exceptional detection power (ppt levels), and the role of modifiers in atomization is the scope for further studies. EcHG offers many advantages, particularly lower costs, freedom from interference, and lower LODs. Despite these, analytical applications reported so far are scarce. The main drawback lies in the production of the reproducible solid cathode surface. Other novel methods of HG include photo-induced generation [290], which is yet to be explored fully.

Elemental speciation based on selective reaction medium continues to be investigated, even though this approach seems to be matured. On the other hand, offline/online SPME techniques and selective HG using a novel reaction medium is a subject of further study, e.g. the use of solid reagents [214]. FI techniques incorporating efficient online preconcentration, matrix modification, and oxidation/reduction for elemental speciation analysis continue to be reported. New approaches to analyte preconcentration techniques such as SPME and SDME have been explored [242]. The use of microorganisms for selective retention of elemental species is an emerging area; however, its applicability to routine analytical applications is scarce.

Sample-pretreatment techniques such as microwave digestion, UV, and US irradiation techniques avoid the use of chemicals and drastic reaction conditions, and allow the determination of non-hydride reactive species (occurring mainly in marine

environments) as well. The photochemical behaviour of organometalloid species offers an alternative route to conventional digestion/prereduction methods.

The field of HG remains a dynamic field of study, as its ability to distinguish between various oxidation states of elements with the use of simple procedures continues to be reported. There is a scope for further improvement if the mechanisms of hydride formation and its atomization in the quartz-tube atomizer are better understood. Recent advances in our understanding of the hydrolysis of THB reagent [291], and discarding of the concept of 'nascent hydrogen' as the mechanism of analyte reduction, may serve to illuminate new approaches to HG, explain the effects of various additives in signal enhancement, and provide a unified approach to optimize the generation of volatile species. The fundamental role played by numerous hydroboran intermediates formed during the derivatization process appears to be vital in these interpretations [292]. The field of HG for elemental speciation analysis is broad and continues to evolve with the development of new methods of generation, e.g. photo-induced HG [290] and preconcentration techniques. Several of the more classical techniques have become stagnant and are used very little for analytical purposes, whereas others are experiencing a resurgence of interest. With the continued improvement in species separation and preconcentration approaches, the HG technique continues to serve as a simple yet powerful tool for the ultra-trace-level determination of elemental species.

#### References

- [1] D.M. Templeton, F. Ariese, R. Cornelis, L.G. Danielsson, H. Muntau, H.P. Van Leeuwen, R. Lobinski. Pure Appl. Chem., 72, 1453 (2000).
- [2] M. Raessler, B. Michalke, S.S. Hostede, A. Kettrup. Sci. Tot. Environ., 258, 171 (2000).
- [3] I. Ipolyi, P. Fodor. Anal. Chim. Acta, 413, 13 (2000).
- [4] T. Nakazato, H. Tao, T. Taniguchi, K. Isshiki. Talanta, 58, 121 (2002).
- [5] E. Terlecka. Environ. Monit. Assess, 107, 259 (2005).
- [6] J.C. Gonzalez, I. Lavilla, C. Bendicho. Talanta, 59, 525 (2003).
- [7] S.L. Anderson, S.A. Pergantis. Talanta, 60, 821 (2003).
- [8] Q. Xie, R. Kerrich, E. Irving, K. Liber, F.A. Shakra. J. Anal. At. Spectrom., 17, 1037 (2002).
- [9] J. Dedina, D.L. Tsalev. Hydride Generation Atomic Absorption Spectroscopy, Wiley, NewYork (1995).
- [10] R. Morabito. Fresenius J. Anal. Chem., 351, 378 (1995).
- [11] I. Havezov. Fresenius J. Anal. Chem., 355, 452 (1996).
- [12] L. Campanella, K. Pyrzynska, M. Trojanowicz. Talanta, 43, 825 (1996).
- [13] F. Zhaolun, T. Guanhong, X. Shukun, L. Xuezhu, W. Jing. Microchem. J., 53, 42 (1996).
- [14] T. Nakahara. Bunseki Kagaku, 46, 513 (1997).
- [15] A.G. Howard. J. Anal. At. Spectrom., 12, 267 (1997).
- [16] D.L. Tsalev. J. Anal. At. Spectrom., 14, 147 (1999).
- [17] Y. Cai. Trends. Anal. Chem., **19**, 62 (2000).
- [18] D.L. Tsalev. Spectrochim. Acta B, 55, 917 (2000).
- [19] A.K. Das, M. de la Guardia, M.L. Cervera. Talanta, 55, 1 (2001).
- [20] P. Niedzielski, M. Siepak, J. Siepak, J. Przybylek. Polish J. Environ. Stud., 11, 219 (2002).
- [21] P. Niedzielski, M. Siepak, J. Przybylek, J. Siepak. Polish J. Environ. Stud., 11, 457 (2002).
- [22] R.E. Sturgeon, Z. Mester. Appl. Spectrosc., 56, 202A (2002).
- [23] M. Burguera, J.L. Burguera. Talanta, 44, 1581 (1997).
- [24] J.H. Weber. Trends Anal. Chem., 16, 73 (1997).
- [25] A.T. de la Calzada, M.C. Villa-Lojo, E. Beceito-Gonzalez, E. Alonso-Rodriguez, D. Prada-Rodriguez. Trends Anal. Chem., 17, 167 (1998).
- [26] C.K. Jain, I. Ali. Wat. Res., 34, 4304 (2000).
- [27] Z. Gong. Talanta, 58, 77 (2002).
- [28] D.G. Kimiburgh, W. Kosmus. Talanta, 58, 165 (2002).
- [29] S. Karthikeyan, S. Hirata. Anal. Lett., 36, 2355 (2003).
- [30] I. Ali, K. Jain. Int. J. Environ. Anal. Chem., 84, 947 (2004).
- [31] D.Q. Hung, O. Nekrassova, R.G. Compton. Talanta, 64, 269 (2004).
- [32] K.A. Francesconi, D. Kuehneit. Analyst, 129, 373 (2004).
- [33] P. Smichowski, Y. Madrid, C. Camara. Fresenius J. Anal. Chem., 360, 623 (1998).
- [34] M.J. Nash, J.E. Maskell, S.J. Hill. *J. Environ. Monit.*, 2, 97 (2000).
- [35] M. Krachler, H. Emons, J. Zheng. Trends Anal. Chem., 20, 79 (2001).
- [36] R.M. Olivas, O.F.X. Donard, C. Camara, P. Quevauviller. Anal. Chim. Acta, 286, 357 (1994).
- [37] X. Dauchy, M. Potin-Gautier, A. Astruc, M. Astruc. *Fresenius J. Anal. Chem.*, 348, 792 (1994).
- [38] K. Pyrzynska. Analyst, 121, 77R (1996).
- [39] A. D'Ulivo. Analyst, 122, 117R (1997).
- [40] K. Pyrzynska. Anal. Sci., 14, 479 (1998).
- [41] K. Pyrzynska. Microchim. Acta, 140, 55 (2002).
- [42] B.D. Wake, A.R. Bowie, E.C.V. Butler, P.R. Haddad. Trends Anal. Chem., 23, 491 (2004).
- [43] J.L. Capelo, C. Fernandez, B. Pedars, P. Santos, P. Gonzalez, C. Vaz. Talanta, 68, 1442 (2006).
- [44] Z. Gong, X. Lu, C. Watt, B. Wen, B. He, J. Mumford, Z. Ning, Y. Xia, X. Chris Le. Anal. Chim. Acta, 555, 181 (2006).
- [45] C.I.S. Narcise, L. dlC Coo, F.R. del Mundo. Talanta, 68, 298 (2005).
- [46] A.N. Anthemidis, G.A. Zachariadis, J.A. Stratis. Anal. Chim. Acta, 547, 237 (2005).
- [47] K. Jitmanee, M. Oshima, S. Motomizu. Talanta, 66, 529 (2005).
- [48] J. Narvaez, P. Richter, M.I. Toral. Anal. Bioanal. Chem., 381, 1483 (2005).
- [49] M.E. Sigrist, H.R. Beldomenico. Spectrochim. Acta B, 59, 1041 (2004).
- [50] X. Yan, X. Yin, X. He, Y. Jiang. Anal. Chem., 74, 2162 (2002).
- [51] S. Maity, S. Chakravarty, P. Thakur, K.K. Gupta, S. Bhattacharjee, B.C. Roy. Chemosphere, 54, 1199 (2004).
- [52] N.M.M. Coelho, A. Cosmen da Silva, C.M. da Silva. Anal. Chim. Acta, 460, 227 (2002).
- [53] J.T. van Elteren, Z. Slejkovec, M. Svetina, A. Glinsek. Fresenius J. Anal. Chem., 370, 408 (2001).
- [54] J.Y. Cabon, N. Cabon. Fresenius J. Anal. Chem., 368, 484 (2000).
- [55] F. El-Hadri, A. Morales-Rubio, M. de la Guardia. Talanta, 52, 653 (2000).
- [56] G. Samanta, T.R. Chowdhury, B.K. Mandal, B.K. Biswas, U.K. Chowdhury, G.K. Basu, C.R. Chanda, D. Lodh, D. Chakraborti. Microchem. J., 62, 174 (1999).
- [57] J.L. Burguera, M. Burguera, C. Rivas, P. Carrero. Talanta, 45, 531 (1998).
- [58] S. Nielsen, E.H. Hansen. Anal. Chim. Acta, 343, 5 (1997).
- [59] D. Schaumloffel, B. Neidhart. Fresenius J. Anal. Chem., 354, 866 (1996).
- [60] J.T. van Elteren, H.A. Das, C.L. de Ligny, J. Agterdenbos, D. Bax. J. Radioanal. Nucl. Chem., 179, 211 (1994).
- [61] S.P. Quinaia, M.C.E. Rollemberg. J. Braz. Chem. Soc., 12, 37 (2001).
- [62] N. Molenat, A. Astruc, M. Holeman, G. Maury, R. Pinel. Analusis, 27, 795 (1999).
- [63] J.M. Bundaleska, T. Stafilov, S. Arpadjan. *Int. J. Environ. Anal. Chem.*, **85**, 199 (2005).
- [64] A. Shraim, B. Chiswell, H. Olszowy. Analyst, 125, 949 (2000).
- [65] K.F. Akter, Z. Chen, L. Smith, D. Davey, R. Naidu. *Talanta*, **68**, 406 (2005).
- [66] P. Niedzielski, M. Siepak. Centrt. Eur. J. Chem., 3, 82 (2005).
- [67] M. Eva Moreno, C. Perez-Conde, C. Camara. J. Anal. At. Spectrom., 15, 681 (2000).
- [68] Y. Chen, M. Zhou, J. Tong, N. Belzile. Anal. Chim. Acta, **545**, 142 (2005).
- [69] X. Tang, Z. Xu, J. Wang. Spectrochim. Acta B, 60, 1580 (2005).
- [70] H. Narasaki, K. Mayumi. Anal. Sci., 16, 65 (2000).
- [71] J.Y. Cabon, T.W. Erler. Analyst, 123, 1565 (1998).
- [72] J. Stripeikis, J. Pedro, A. Bonivardi, M. Tudino. Anal. Chim. Acta, 502, 99 (2004).
- [73] M.G. Cobo Fernandez, M.A. Palacios, C. Camara. Anal. Chim. Acta, 283, 386 (1993).
- [74] K. Itoh, M. Chikuma, M. Nishimura, T. Tanaka, M. Tanaka, M. Nakayama, H. Tanaka. Fresenius Z Anal. Chem., 333, 102 (1989).
- [75] U. Ornemark, A. Olin. Talanta, 41, 67 (1994).
- [76] A. Larraya, M.G. Cobo-Fernandez, M.A. Palacios, C. Camara. Fresenius J. Anal. Chem., 350, 667 (1994).
- [77] K.J. Reddy, Z. Zhang, M.J. Blaylock, G.F. Vance. *Environ. Sci. Tech.*, 29, 1754 (1995).
- [78] C. Brunori, M.B. de la Calle-Guntinas, R. Morabito. Fresenius J. Anal. Chem., 360, 26 (1998).
- [79] G.A. Cutter. Anal. Chim. Acta, 98, 59 (1978).
- [80] S. Garbos, E. Bulska, A. Hulanicki, N.I. Shcherbinina, E.M. Sedykh. Anal. Chim. Acta, 342, 167 (1997).
- [81] J.Y. Cabon, C.L. Madec. Anal. Chim. Acta, 504, 209 (2004).
- [82] S. Garbos, M. Rzepecka, E. Bulska, A. Hulanicki. Spectrochim. Acta B, 54, 873 (1999).
- [83] T. Deng, Y. Chen, N. Belzile. Anal. Chim. Acta, 432, 293 (2001).
- [84] P. Niedzielski, M. Siepak, K. Grabowski. Polish J. Environ. Stud., 12, 213 (2003).
- [85] M.O. Yu, G.O. Liu, O. Jin. Talanta, 30, 265 (1983).
- [86] P. Niedzielski, M. Siepak. Anal. Lett., 36, 971 (2003).
- [87] R.S. Braman, C.C. Foreback. Science, 182, 1247 (1973).
- [88] B.K. Mandal, K.I. Suzuki. Talanta, 58, 201 (2002).
- [89] A.H. Smith, P.A. Lopipero, M.N. Bates, C.M. Steinmaus. Science, 296, 2145 (2002).
- [90] J. Zhu, Z. Chen, V. Lallemand-Breitenbach, H. de The. Nat. Rev. Cancer, 2, 705 (2002).
- [91] J. Aggett, A.C. Aspell. Analyst, 101, 341 (1976).
- [92] S. Nakashima. Analyst, 104, 172 (1979).
- [93] T. Nakahara. Anal. Chim. Acta, 131, 73 (1981).
- [94] W.A. Maher. Anal. Chim. Acta, 126, 157 (1981).
- [95] K. Terada, K. Matsumoto, T. Inaba. Anal. Chim. Acta, 158, 207 (1984).
- [96] A.G. Howard, M.H. Arbab-Zavar. Analyst, 105, 338 (1980).
- [97] J.S. Edmond, K.A. Francesconi. Anal. Chem., 48, 2019 (1976).
- [98] T.A. Hinners. Analyst, 105, 751 (1980).
- [99] A. Lopez, R. Torralba, M.A. Palacios, C. Camara. Talanta, 39, 1343 (1992).
- [100] A.J. Narsito, J. Agterdenbos. Anal. Chim. Acta, 197, 315 (1987).
- [101] M. Yamamoto, K. Urata, K. Murashige, Y. Yamamoto. Spectrochim. Acta, 36 B, 671 (1981).
- [102] M. Yamamoto, K. Fujishige, H. Tsubota, Y. Yamamoto. Anal. Sci., 1, 47 (1985).
- [103] M. Yamamoto, M. Yasuda, Y. Yamamoto. Anal. Chem., 57, 1382 (1985).
- [104] R.B. McCleskey, D.K. Nordstrom, A.S. Maest. Appl. Geochem., 19, 995 (2004).
- [105] R.K. Anderson, M. Thomson, E. Culbard. Analyst, 111, 1143 (1986).
- [106] I.D. Brindle, H. Alarabi, S. Karshman, X.-C. Le, S. Zheng, H. Chen. Analyst, 117, 407 (1992).
- [107] H. Chen, I.D. Brindle, X. Lee. Anal. Chem., 64, 667 (1992).
- [108] C. Boampong, I.D. Brindle, X.-C. Lee, L. Pidwerbesky, C.M.C. Ponzoni. Anal. Chem., 60, 1185 (1988).
- [109] P. Niedzielski, J. Siepak, Z. Kowalczuk. Polish J. Environ. Stud., 8, 183 (1999).
- [110] P. Liang, A. Li. Fresenius J. Anal. Chem., 368, 418 (2000).
- [111] P. Niedzielski. *Environ. Monitt. Assess*, **118**, 231 (2006).
- [112] R. Torralba, M. Bonilla, L.V. Perez-Arribas, A. Palacios, C. Camara. Spectrochim. Acta B, 49, 893 (1994).
- [113] R. Torralba, M. Bonilla, L.V. Perez-Arribas, M.A. Palacios, C. Camara. Microchim. Acta, 126, 257 (1997).
- [114] Y. Yano, T. Miyama, A. Ho, T. Yasuda. Anal. Sci., 16, 939 (2000).
- [115] T.R. Rude, H. Puchelt. Fresenius J. Anal. Chem., 350, 44 (1994).
- [116] S.P. Quinaia, M.C.E. Rollemberg. *J. Braz. Chem. Soc.*, 8, 349 (1997).
- [117] A. Shraim, B. Chiswell, H. Olszowy. Talanta, **50**, 1109 (1999).
- [118] M. Borho, P. Wilderer. J. Wat. Suppl. Res. Technol.-AQUA, 46, 138 (1997).
- [119] J. Muller. Fresenius J. Anal. Chem., 363, 572 (1999).
- [120] C.H. Chung, E. Iwamoto, M. Yamamoto, Y. Yamamoto. Spectrochim. Acta, 39B, 459 (1984).
- [121] W.M. Mok, C.M. Wai. Anal. Chem., 59, 233 (1987).
- [122] W.M. Mok, J.A. Riley, C.M. Wai. Wat. Res., 22, 769 (1988).
- [123] S.A. Amankwah, J.L. Fasching. Talanta, 32, 111 (1985).
- [124] K.S. Subramanian, J.C. Meranger. Anal. Chim. Acta, 124, 131 (1981).
- [125] D. Chakraborti, F. Adams, K.J. Irgolic. Fresenius Z Anal. Chem., 323, 340 (1986).
- [126] G. Bombach, W. Klemm, A. Greif. Microchim. Acta, 151, 203 (2005).
- [127] M. Okumura, K. Fujinaca, Y. Seike, M. Nagata, S. Matsuo. Bunseki Kagaku, 52, 1147 (2003).
- [128] A. Chatterjee, D. Das, B.K. Mandal, T.R. Chowdhury, G. Samanta, D. Chakraborti. Analyst, 120, 643 (1995).
- [129] A.R.K. Dapaah, A. Ayame. Anal. Sci., 13, 405 (1997).
- [130] J.T. van Elteren, N.G. Haselager, H.A. Das, C.L. de Lingy, J. Agterdenbos. Anal. Chim. Acta, 252, 89 (1991).
- [131] R.S. Braman, D.L. Johnson, C.C. Foreback, J.M. Ammons, J.L. Bricker. Anal. Chem., 49, 821 (1977).
- [132] C. Feldman. Anal. Chem., **51**, 664 (1979).
- [133] M.O. Andreae. Anal. Chem., 49, 820 (1977).
- [134] A.G. Howard, M.H. Arbab-Zavar. Analyst, 106, 213 (1981).
- [135] R.J.A. Van Cleuvenbergen, W.E. Van Mol, F.C. Adams. J. Anal. At. Spectrom., 3, 169 (1988).
- [136] R.K. Anderson, M. Thomson, E. Culbard. Analyst, 111, 1153 (1986).
- [137] J. Ryu, R.A. Dahlgren, R.A. Zierenberg. Geochem. Cosmochim. Acta, 66, 2981 (2002).
- [138] A.G. Howard, S.D.W. Comber. *Microchim. Acta*, 109, 27 (1992).
- [139] P. Michel, B. Averty, V. Colandini. Microchim. Acta, 109, 35 (1992).
- [140] M. Burguera, J.L. Burguera, M.R. Brunetto, M. de la Guardia, A. Salvandar. Anal. Chim. Acta, 261, 105 (1992).
- [141] A.G. Howard, C. Salou. Anal. Chim. Acta, 333, 89 (1996).
- [142] A.G. Howard, C. Salou. J. Anal. At. Spectrom., 13, 683 (1998).
- [143] H. Hasegawa, Y. Sohrin, M. Matsui, M. Hojo, M. Kawashima. Anal. Chem., 66, 3247 (1994).
- [144] H. Hasegawa, M. Matsui, S. Okamura, M. Hojo, N. Iwasaki, Y. Sohrin. Appl. Organomet. Chem., 13, 113 (1999).
- [145] J.Y. Cabon, N. Cabon. Anal. Chim. Acta, 418, 19 (2000).
- [146] T.M. Hsiung, J.M. Wang. J. Anal. At. Spectrom., 19, 923 (2004).
- [147] O. Astrom. Anal. Chem., **54**, 190 (1982).
- [148] P.K. Hon, O.W. Lau, S.K. Tusi. *J. Anal. At. Spectrom.*, 1, 125 (1986).
- [149] W. Driehaus, M. Jekel. Fresenius J. Anal. Chem., 343, 352 (1992).
- [150] S.P. Schwenzer, C.E.T. Michael Kersten, T. Kirnhaver. Fresenius J. Anal. Chem., 371, 927 (2001).
- [151] B. Welz, M. Sucmanova. Analyst, 118, 1417 (1993).
- [152] B. Welz, M. Sucmanova. Analyst, 118, 1425 (1993).
- [153] X. Yin, E. Hoffmann, C. Ludke. *Fresenius J. Anal. Chem.*, 355, 324 (1996).
- [154] E.A. Cordos, T. Frentiu, M. Ponta, B. Abraham. Chem. Speci. Bioavail., 18, 1 (2006).
- [155] J.F. Tyson, S.G. Offky, N.J. Seare, H.A. Kibble, C. Fellows. J. Anal. At. Spectrom., 7, 315 (1992).
- [156] S.-K. Xu, Z.L. Fang. Fenxi Shiyanshi, 13, 20 (1994).
- [157] X.C. Lee, W.R. Cullen, K.J. Reimer. Anal. Chim. Acta, 285, 277 (1994).
- [158] P. Carrero, A. Malave, J.L. Burguera, M. Burguera, C. Rondo. Anal. Chim. Acta, 438, 195 (2001).
- [159] R. Torralba, M. Bonilla, A. Placios, C. Camara. Analusis, 22, 478 (1994).
- [160] S. Nielsen, J.J. Sloth, E.H. Hansen. Talanta, 43, 867 (1996).
- [161] G.G. Bortoleto, S. Cadore. Talanta, 67, 169 (2005).
- [162] S. Karthikeyan, S. Hirata. Anal. Bioanal. Chem., 375, 139 (2003).
- [163] Y. Feng, J. Cao. Anal. Chim. Acta, 293, 211 (1994).
- [164] E.J. Knudson, G.D. Christian. At. Absorpt. Newsl., 13, 74 (1974).
- [165] H. Matusiewicz, R.E. Sturgeon. Spectrochim. Acta B, 51, 377 (1996).
- [166] R. Kalahne, G. Henrion, A. Hulanicki, S. Garbos, M. Walcerz. Spectrochim. Acta B, 52, 1509 (1997).
- [167] R.E. Sturgeon, S.N. Wille, S.S. Berman. Fresenius Z Anal. Chem., 323, 788 (1986).
- [168] X.-P. Yan, Z.-M. Ni. Anal. Chim. Acta, 291, 89 (1994).
- [169] Z. Fang, G. Tao. Fresenius J. Anal. Chem., 355, 576 (1996).
- [170] A.K. Das, R. Chakraborty. Fresenius J. Anal. Chem., 357, 1 (1997).
- [171] A.U. Shaikh, D.E. Tallman. Anal. Chim. Acta, 98, 251 (1978).
- [172] P. Niedzielski, M. Siepak. Chem. Ecol., 21, 241 (2005).
- [173] K. Anezaki, I. Nukatsuka, K. Ohzeki. Anal. Sci., 15, 829 (1999).
- [174] C. Hsieh, C. Yen, M. Kuo. Anal. Sci., 15, 669 (1999).
- [175] R. Murata, T. Shimizu, N. Uehara. Bunseki Kagaku, 54, 831 (2005).
- [176] E. Ruseva, I. Havezov, A. Detcheva. Fresenius J. Anal. Chem., 347, 320 (1993).
- [177] P. Bermejo-Barrera, J. Moreda-Pineiro, A. Moreda-Pineiro, A. Bermejo-Barrera. Anal. Chim. Acta, 374, 231 (1998).
- [178] S.N. Willie. Spectrochim. Acta B, 51, 1781 (1996).
- [179] R.E. Sturgeon, K.W. Michael Siu, S.N. Wille, S.S. Berman. Analyst, 114, 1393 (1989).
- [180] P.H. Hermelo, M.C.B. Alonso, A.B. Barrera, P.B. Barrera. J. Anal. At. Spectrom., 20, 662 (2005).
- [181] M. Morita, J.J. Edmonds. Pure Appl. Chem., 64, 575 (1992).
- [182] V.I. Salveykova, F. Rastegar, M.J.F. Leroy. J. Anal. At. Spectrom., 11, 997 (1996).
- [183] P. Niedzielski, M. Siepak, J. Siepak. *Microchem. J.*, **72**, 137 (2002).
- [184] I.K. Tsujii, K. Kuga. Anal. Chim. Acta, 72, 85 (1974).
- [185] I.K. Tsujii, K. Kuga. Anal. Chim. Acta, 97, 51 (1978).
- [186] Z. Mester, P. Fodor. Spectrochim. Acta B, 52, 1763 (1997).
- [187] A. D'Ulivo, E. Bramanti, L. Lampugnani, R. Zamboni. Spectrochim. Acta B, 56, 1893 (2001).
- [188] X.C. Lee, S. Yalcin, M. Ma. *Environ. Sci. Tech.*, 34, 2342 (2000).
- [189] A.C. Lopez, M.D.L. de Castro. *J. Anal. At. Spectrom.*, **17**, 1363 (2002).
- [190] E. Castro, I. Lavilla, C. Bendicho. Talanta (In press).
- [191] A.M. Featherstone, E.C.V. Butler, B.V. O'Grandy, P. Michel. J. Anal. At. Spectrom., 13, 1355 (1998).
- [192] L.O. Leal, R. Forteza, V. Cerda. Talanta, 69, 500 (2006).
- [193] E. Denkhaus, A. Golloch, X.-M. Guo, B. Huang. J. Anal. At. Spectrom., 16, 870 (2001).
- [194] E. Denkhaus, F. Beck, P. Bueschler, R. Gerhard, A. Golloch. Fresenius J. Anal. Chem., 370, 735 (2001).
- [195] W.W. Ding, R.E. Sturgeon. Spectrochim. Acta B, 51, 1325 (1996).
- [196] D. Schaumloffel, B. Neidhart. Fresenius J. Anal. Chem., 354, 866 (1996).
- [197] U. Pyell, A. Dworschak, F. Nitschke, B. Neidhart. Fresenius J. Anal. Chem., 363, 495 (1999).
- [198] X. Li, J. Jia, Z. Wang. Anal. Chim. Acta, 560, 153 (2006).
- [199] Y. Madrid, C. Camara. Trends Anal. Chem., 16, 36 (1997).
- [200] B. Godlewska-Zylkiewicz. Crit. Rev. Anal. Chem., 31, 175 (2001).
- [201] P. Smichowski, J. Marrero, A. Ledesma, G. Polla, D.A. Batistoni. J. Anal. At. Spectrom., 15, 1493 (2000).
- [202] J. Koh, Y. Kwon, Y. Pak. Microchem. J., 80, 195 (2005).
- [203] W.T. Frankenberger, S. Benson. Selenium in the Environment, Dekker, New York (1994).
- [204] M.E. Moreno, C. Perez-Conde, C. Camara. Anal. Bioanal. Chem., 375, 666 (2003).
- [205] P.N. Vijan, D. Leung. Anal. Chim. Acta, 120, 141 (1980).
- [206] J.P. Diaz, M. Navarro, H. Lopez, M.C. Lopez. Sci. Tot. Environ., 186, 231 (1996).
- [207] S.C. Apte, A.G. Howard. J. Anal. At. Spectrom., 1, 379 (1986).
- [208] I.D. Gregori, M.G. Lobos, H. Pinochet. Wat. Res., 36, 115 (2002).
- [209] F. Nakata, Y. Yasui, H. Matsuo, T. Kumamaru. Anal. Sci., 1, 417 (1985).
- [210] Y. Tamari, H. Ogura. Bunseki Kagaku, 46, 313 (1997).
- [211] C.C. Johnson, X. Ge, K.A. Green, X. Liu. Appl. Geochem., 15, 385 (2000).
- [212] G.A. Cutter. Science, 217, 829 (1982).
- [213] E.M.M. Flores, S.R. Mortari, A.F. Martins. J. Anal. At. Spectrom., 12, 379 (1997).
- [214] N. Maleki, A. Safavi, M.M. Doroodmand. Talanta, 66, 858 (2005).
- [215] J. Stripeikis, M. Tudino, O. Troccoli, R. Wuillound, R. Olsina, L. Martinez. Spectrochim. Acta B, 56, 93 (2001).
- [216] S. Schermer, L. Jurica, J. Paumard, E. Beinrohr, F.M. Matysik, J.A.C. Broekaert. Fresenius J. Anal. Chem., 371, 740 (2001).
- [217] Y. Tamari, H. Ogura. Bunseki Kagaku, 46, 605 (1997).
- [218] K.J. Lamble, S.J. Hill. Analyst, 123, 103R (1998).
- [219] G. Tao, E.H. Hansen. Analyst, 119, 333 (1994).
- [220] R. Rubio, A. Padro, C. Rauret. Anal. Chim. Acta, 353, 91 (1997).
- [221] L. Pitts, P.J. Worsfold, S.J. Hill. Analyst, 119, 2785 (1994).
- [222] D.W. Bryce, A. Izquierdo, M.D.L. de Castro. J. Anal. At. Spectrom., 10, 1059 (1995).
- [223] R.M. Olivas, O.F.X. Donard. Talanta, 45, 1023 (1998).
- [224] Z. Wang, Y. Gao, N. Belzile. Anal. Chem., 73, 4711 (2001).
- [225] J.L. Burguera, P. Carrero, M. Burguera, C. Rondon, M.R. Brunetto, M. Gallignani. Spectrochim. Acta B, 51, 1837 (1996).
- [226] J.M.G. LaFuente, M.L.F. Sanchez, J.M. Marchante-Gayon, J.E.S. Uria, A. Sanz-Medel. Spectrochim. Acta B, 51, 1849 (1996).
- [227] M. Gallignaani, M. Valero, M.R. Brunetto, J.L. Burguera, M. Burguera, Y.P. de Pena. Talanta, 52, 1015 (2000).
- [228] H. Mendez, I. Lavilla, C. Bendicho. J. Anal. At. Spectrom., 19, 1379 (2004).
- [229] C. Harzdorf, G. Janser, D. Rinne, M. Rogge. Anal. Chim. Acta, 374, 209 (1998).
- [230] J. Stripeikis, P. Costa, M. Tudino, O. Troccoli. Anal. Chim. Acta, 408, 191 (2000).
- [231] K. Pyrzynska. Analyst, 120, 1933 (1995).
- [232] K. Pyrzynska, P. Drzemyslaw, M. Trojanowicz. Anal. Chim. Acta, 363, 141 (1998).
- [233] P.E. Carrero, J.F. Tyson. Analyst, 122, 915 (1997).
- [234] U. Ornemark, A. Olin. Talanta, 41, 1675 (1994).
- [235] J.L. Gomez-Ariza, J.A. Pozas, I. Giraldez, E. Morales. Analyst, 124, 75 (1999).
- [236] F. Sahin, M. Volkan, A.G. Howard, O.Y. Ataman. Talanta, 60, 1003 (2003).
- [237] D.R. Rodden, D.E. Tallman. Anal. Chem., 54, 307 (1982).
- [238] S. Li, N. Deng. Anal. Bioanal. Chem., 374, 1341 (2002).
- [239] K.O. Saygi, E. Melek, M. Tuzen, M. Soylak. Talanta (In press).
- [240] P. Niedzielski, M. Siepak, B. Dudzinska-Huczuk. Centrt. Eur. J. Chem., 3, 314 (2003).
- [241] F. Sahin, M. Volkan, O.Y. Ataman. Anal. Chim. Acta, 547, 126 (2005).
- [242] S. Fragueiro, I. Lavilla, C. Bendicho. Talanta, 68, 1096 (2006).
- [243] M. Tuzen, K.O. Saygi, M. Soylak. Talanta (In press).
- [244] Y. He, J. Moreda-Pineiro, M.L. Cervera, M. de la Guardia. J. Anal. At. Spectrom., 13, 289 (1998).
- [245] C.-Y. Lu, X.-P. Yan, Z.-P. Zhang, Z.-P. Wang, L.-W. Liu. J. Anal. At. Spectrom., 19, 277 (2004).
- [246] Y.-W. Chen, X.-L. Zhou, J. Tong, Y. Truong, N. Belzile. Anal. Chim. Acta, 545, 149 (2005).
- [247] T. Perez-Corona, Y. Madrid, C. Camara. Anal. Chim. Acta, 345, 249 (1997).
- [248] L.C. Robles, J.C. Feo, B. de Celis, J.M. Lumbreras, C. Garcia-Olalla, A.J. Aller. Talanta, 50, 307 (1999).
- [249] J. Piwonka, G. Kaiser, G. Tolg. Fresenius Z Anal. Chem., 321, 225 (1985).
- [250] R. Bye. Talanta, 30, 993 (1983).
- [251] V. Krivan, K. Petrick, B. Welz, M. Melcher. Anal. Chem., 57, 1703 (1985).
- [252] S.P. Brimmer, W.R. Fawcett, K.A. Kulhavy. Anal. Chem., 59, 1470 (1987).
- [253] S.J. Hill, L. Pitts, P. Worsfold. J. Anal. At. Spectrom., 10, 409 (1995).
- [254] R. Bye, W. Lund. Fresenius Z Anal. Chem., 332, 242 (1988).
- [255] J. Petterson, A. Olin. Talanta, 38, 413 (1991).
- [256] A. D'Ulivo. *J. Anal. At. Spectrom.*, **4**, 67 (1989).
- [257] A. Ramesh Kumar, P. Riyazuddin. Microchim. Acta, 155, 387 (2006).
- [258] I.D. Brindle, E. Lugowska. Spectrochim. Acta B, 52, 163 (1997).
- [259] A. D'Ulivo, L. Lampugnani, I. Sfetsios, R. Zamboni, C. Forte. Analyst, 119, 633 (1994).
- [260] N.O. Kenduzler, G. Somer, P. Zuman. Talanta, 69, 25 (2006).
- [261] W. Shotyk, M. Krachler, B. Chen. J. Environ. Monit., 7, 1135 (2005).
- [262] S. Nakashima. Analyst, 105, 732 (1980).
- [263] L.S. Cutter, G.A. Cutter, M.L.C. San Diego-Mc Glone. Anal. Chem., 63, 1138 (1991).
- [264] S.C. Apte, A.G. Howard. *J. Anal. At. Spectrom.*, 1, 221 (1986).
- [265] M.O. Andreae, J.F. Asmode, P. Foster, L. Vant Dack. Anal. Chem., 53, 1766 (1982).
- [266] A.T. Campbell, A. Howard. Anal. Proc., 26, 32 (1989).

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- [267] M.B. de la Calle-Guntinas, Y. Madrid, C. Camara. Fresenius J. Anal. Chem., 343, 597 (1992).
- [268] T. Nakahara, N. Kikui. Anal. Chim. Acta, 172, 127 (1985).
- [269] H.B. Hou, H. Narasaki. Anal. Sci., 14, 1161 (1998).
- [270] M.B. de la Calle-Guntinas, Y. Madrid, C. Camara. Microchim. Acta, 109, 149 (1992).
- [271] A.M. Gracia, M.C.P. Rodriguez, J.E.S. Uria, A. Sanz-Medal. *Fresenius J. Anal. Chem.*, 353, 128 (1995).
- [272] Y. Feng, H. Narasaki, H. Chen, L. Tian. Anal. Chim. Acta, 386, 297 (1999).
- [273] N. Ulrich. Anal. Chim. Acta, 417, 201 (2000).
- [274] J.R. Castillo, C. Martinz, P. Chamorro, J.M. Mir. Microchim. Acta, III, 95 (1986).
- [275] B. de la Calle-Guntinas, Y. Madrid, C. Camara. Anal. Chim. Acta, 247, 7 (1991).
- [276] H. Fukuda, J. Tsunoda, K. Matsumoto, K. Kerada. Bunseki Kagaku, 36, 683 (1987).
- [277] B. de la Calle-Guntinas, Y. Madrid, C. Camara. Anal. Chim. Acta, 252, 161 (1991).
- [278] A. Erdem, A.E. Eroglu. Talanta, 68, 86 (2005).
- [279] F.-y. Zheng, S.-y. Qian, S.-x. Li, X.-Q. Huang, L.-x. Lin. Anal. Sci., 22, 1319 (2006).
- [280] P. Smichowski, B. de la Calle-Guntinas, Y. Madrid, M.G. Cobo, C. Camara. Spectrochim. Acta, 49B, 1049 (1994).
- [281] A. D'Ulivo, L. Lampugnani, G. Pellegrini, R. Zamboni. J. Anal. At. Spectrom., 10, 969 (1995).
- [282] K. Tsujii. Anal. Lett., 14, 181 (1981).
- [283] Y.R. Suo, Y.L. Huang. Guangpuxue Yu Guangpu Fenxi, 12, 87 (1992).
- [284] T. Kamada, N. Sugita, Y. Yamamoto. Talanta, 26, 337 (1979).
- [285] I. Havezov, N. Jordanov. Talanta, 21, 1013 (1974).
- [286] M.O. Andreae. Anal. Chem., 56, 2064 (1984).
- [287] D.S. Lee, J.S. Edmond. Nature, 313, 782 (1985).
- [288] M.M. Yoon, S.C. Shim, H.C. Pyun, D.S. Lee. Anal. Sci., 6, 561 (1990).
- [289] A. Korez, A.E. Eroglu, M. Volkan, O.Y. Ataman. J. Anal. At. Spectrom., 15, 1599 (2000).
- [290] X. Guo, R.E. Sturgeon, Z. Mester, G.J. Gardner. Anal. Chem., 75, 2092 (2003).
- [291] A. D'Ulivo. Spectrochim. Acta B, 59, 793 (2004).
- [292] A. D'Ulivo, Z. Mester, R.E. Sturgeon. Spectrochim. Acta B, 60, 423 (2005).